

Rossella Crescitelli, PhD - Curriculum vitae

- **PERSONAL INFORMATION**

NAME Rossella Crescitelli
BIRTH 7 March 1985
PHONE + 46 (0)72 873 15 19
E-MAIL rossella.crescitelli@gu.se
FAMILY Married, two children
RERERENTS Prof. Roger Olofsson Bagge, Sahlgrenska University Hospital, University of Gothenburg
Prof. Jan Lötvall, Sahlgrenska Akademy, University of Gothenburg

- **CURRENT POSITION**

2024 - **Junio PI**
Sahlgrenska Center for Cancer Research, University of Gothenburg
Research topic: Investigation of extracellular vesicles in the tumor microenvironment

- **PREVIOUS WORK EXPERIENCES**

2020 - 2024 **Senior Researcher**
Sahlgrenska Center for Cancer Research, University of Gothenburg
Research topic: Investigation of the role of uveal melanoma-derived extracellular vesicles in the formation of pre-metastatic niches in the liver.

2014 - 2019 **Post doc fellow**
Krefting Research Centre, University of Gothenburg
Research topic: Isolation and characterization of extracellular vesicles from tissues

2021 **Guest researcher**
University of Groningen, Groningen, Netherland
Prof: Peter Olinga

2016 **Guest researcher**
Umeå University, Umeå Core Facility for Electron Microscopy (UCEM)
Facility leader: Linda Sandblad

2015 **Guest researcher**
Pohang University of Science and Technology, Pohang, Republic of Korea
Prof. Jong Song Gho

2012 - 2013 **Guest PhD Student**
Krefting Research Centre, University of Gothenburg
Research topic: Extracellular vesicles: development of isolation methods

2007 - 2010 **Intern**
Dept. of Biochemistry and Medical Biotechnology,
University of Naples “Federico II”
Research topic: Gene transcriptional analysis: anti-apoptotic gene bag3 as a new target gene of transcription factor WT1

2006 - 2007 **Intern**
Dept. of Clinical Pathology – Functional area of cytogenetics in prenatal diagnosis,
University of Naples “Federico II”

- **EDUCATION**

MAY 2024 **Associate Professorship in Cell and Molecular Biology**

APR 2014 **PhD in Biotechnology for Human Health**
Dept. of Medical Sciences, University of Eastern Piedmont “Amedeo Avogadro”
Research topic: Extracellular vesicle analysis in erythroid ribosomal stress (Diamond Blackfan Anemia)

MAR 2010 **Master degree in Medical Biotechnology**
University of Naples “Federico II”

OCT 2007 **Bachelor degree in Medical Biotechnology**
University of Naples “Federico II”

- **TEACHING ACTIVITIES (TOTAL 212 H)**

- NOV2024 **Lecturer** (SK00134) at University of Gothenburg Extracellular vesicles: isolation from bacteria and therapeutic usage”
- NOV 2023 **Coordinator of PhD course** (SK00034) at University of Gothenburg “Extracellular vesicles: from biology to isolation from tissues”
- MAY 2023 **Lecturer:** ISEV2023 annual meeting, education day “EV characterization methods: practical tips and tricks”
- JUNE 2022 **Guest lecturer**
10 hours of lectures about extracellular vesicles with focus on Cancer for PhD course in Experimental Oncology and Surgery of the University of Palermo, Italy and Graduate School in Oncology University of Palermo, Italy
- JUNE 2022 **Guest lecturer**
10 hours of lectures about extracellular vesicles with focus on neurodegenerative diseases for PhD course in Biomedicine, Neuroscience and Advanced diagnostics of the University of Palermo, Italy
- OCT 2022 University of Gothenburg - PhD course in liquid biopsy
- OCT 2022 University of Naples - Erasmus Mundus program in BeInPrecision Medicine (BeinPM)
- OCT 2022 University of Eastern Piedmont “Amedeo Avogadro”, Novara, Italy
Master’s degree in Medicine and Surgery
Master Degree in Medical Biotechnologies in applied immunology in tissue regeneration
Master Degree in Medical Biotechnologies in transplant immunology
Bachelor Degree in Medical Laboratory Science
PhD program in Sciences and Medical Biotechnologies and in Food, Health and Longevity
- OCT 2022 University ‘G. d’Annunzio’ of Chieti-Pescara (Italy) - PhD course
- NOV 2022 Lecture for Surgeon about extracellular vesicles and cancer
- 2010 - 2012 **Teacher assistant**
100 hours of tutorship and supplementary teaching activity for Department of Translational Medicine and faculty of Mathematic, Physical, Natural Sciences, University of Eastern Piedmont “Amedeo Avogadro”, Novara, Italy

- **REVIEWING ACTIVITIES**

- Referee for journal:** 14 journals from publisher like Journal of Extracellular Vesicles, Nature Communications and Respiratory Research.
- Editor for scientific journal:** guest editor for a special issue in Frontiers in Genetics “The Role of Extracellular Vesicles in Diseases: shedding light on their role as cell-to-cell communication” (2021-2022)
- Poster judge** (ISEV conference 2021, 2022, 2023)
- Abstract judge** (ISEV conference 2023, 2024)
- External assessor**, PhD thesis committee at University of Brescia, Italy (2023), University of G. D’Annunzio, Pescara, Italy (2024)
- Co-chair of oral session** (ISEV conference 2023, MOVE conference 2024)

- **CAREER BREAKS (TOTAL: 24 MONTHS)**

- 2017 Full-time parental leave, 9 months
- 2017 - 2018 Part-time (50 %) parental leave, 7 months
- 2019 Full-time parental leave, 9 months
- 2022 Part-time (20 %) parental leave, 3 months

- **AWARDS**

- 2025 Project-bound PhD position (Sahlgrenska Academy)
- 2024 Top Senior Interpretative Competition Award – ISEV 2024
- 2023, 2022, 2019, 2015 Traveling award: Stiftelsen Assar Gabrielssons Fond.
- 2023, 2022 Traveling award: Cancer Fonden
- 2020 Scholarship ISEV 2020

- **PATENT**

Tissue-Derived Extracellular Vesicles and Their Use as Diagnostics, Registration number:

US20200088734A1, Date of registration: 2017-03-23

Bacteria-derived vesicles and uses thereof, Registration number: WO2020146390A1, Date of registration: 2019-01-09

- **COMMUNICATIONS**

Submitted more than thirty abstracts to international scientific meetings. Some of them chosen for oral communication (MOVE 2024, ISEV 2021 and 2023, 2nd EVIta symposium (2021), Euro-BioImaging User Forum (2021), FAONS – ICN (2022)

Invited speaker to eleven seminars among them: WebEVTalk, TechEM Seminar, SNEV talk, cONCRete - "Cancer Exosomes: small players, big role", Vanderbilt Center for Extracellular Vesicle Research, MIVAC, University of Eastern Piedmont “Amedeo Avogadro” and symposiums

- **PEDAGOGICAL COURSES**

2024 **PIL102** Teaching and Learning in Higher Education 2 (University of Gothenburg)

2023 **PIL101** Teaching and Learning in Higher Education 1 (University of Gothenburg)

2022 **PIL201** Supervision in Postgraduate Programmes (University of Gothenburg)

- **COURSES**

2018 Workshop in electron tomography, University of Gothenburg (Sweden)

2016 Licence in **animal work** (mouse and rat), University of Gothenburg (Sweden)

2015 Licence in “Hårdplasthantering” for resin manipulation for EM sample preparation, University of Gothenburg (Sweden)

2014 Courses on **Nanosight** instrumentation, **CyFlow[®] Cube 6** (Partec), **qNano** (IZON), **ZetaView[®]** (Particle Matrix), **BD FACSVerser**, University of Gothenburg (Sweden)

2012 Course on **BD FACSCalibur** instrumentation, Bekton Dickinson Company, Buccinasco (Italy)

- **TECHNICAL SKILLS**

Cell biology techniques

Bacterial and Mammalian cell cultures, Stem cell cultures (CD34+ cells extraction from cordon blood, mesenchymal stem cells), Cell transfection (transient and stable) by liposome technique and electroporation, Cell transduction by lentivirus vectors, Flow cytometry, Extracellular vesicle isolation from different tissues and biological liquids by different methods (differential centrifugation, density gradient ultracentrifugation antibody-coated magnetic beads, size exclusion chromatography, commercial kits), animal (mouse) work.

Biochemistry and molecular biology techniques

Nucleic acid extraction from cells and vesicles, Electrophoretic techniques, Reverse transcriptase-polymerase chain reaction (RT-PCR), Real Time-PCR (TaqMan, Syber Green), Cloning techniques, Luciferase assay, Extraction protein techniques, Western blotting, Production of recombinant proteins, Chromatin immunoprecipitation (CHIP), Co-immunoprecipitation, Viability assays (BrdU, MTT)

Cytogenetics and prenatal diagnostic techniques

Amniocyte cultures, Reconstruction of karyotypes by the use of karyotyping, Fluorescence in situ hybridization (FISH)

Imaging techniques

Sample preparation for negative stain

Tissue preparation: high pressure freezing, freeze substitution and chemical fixation

Imaging acquisition:

- transmission electron microscopy (Talos L120C, LEO 912 OMEGA and JEOL 1230)
- scanning electron microscopy (ZEISS GeminiSEM 450)
- light microscopy (Axio Observer and cellR/scanR)

Imaging reconstruction: electron tomography

Imaging analysis: IMOD software

- **LAB SOFTWARE**

Image reconstruction (IMOD), FlowJo, 2100 Bioanalyzer software, Real Time PCR data analysis (Life Technologies, Applied Biosystem)

- **LANGUAGE SKILLS**

ITALIAN Mother tongue

ENGLISH Advanced

SWEDISH Beginner